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Spastic Paraplegia 7

Synonym: Hereditary Spastic Paraplegia, Paraplegin Type Giorgio Casari, PhD^{1,2} and Roberto Marconi, MD³ Created: August 24, 2006; Updated: October 25, 2018.

Summary

Clinical characteristics

Spastic paraplegia 7 (SPG7) is characterized by insidiously progressive bilateral leg weakness and spasticity. Most affected individuals have decreased vibration sense and cerebellar signs. Onset is mostly in adulthood, although symptoms may start as early as age 11 years and as late as age 72 years. Additional features including ataxia (gait and limbs), spastic dysarthria, dysphagia, pale optic disks, ataxia, nystagmus, strabismus, ptosis, hearing loss, motor and sensory neuropathy, amyotrophy, scoliosis, *pes cavus*, and urinary sphincter disturbances may be observed.

Diagnosis/testing

The diagnosis of SPG7 is established in a proband with typical clinical findings and biallelic pathogenic variants in *SPG7* identified by molecular genetic testing.

Management

Treatment of manifestations: Drugs that may reduce spasticity and muscle tightness include baclofen, tizanidine, dantrolene, and diazepam. Physical therapy and assistive walking devices often reduce contractures, provide support, and promote stability. Occupational therapy and speech therapy help with activities of daily living.

Surveillance: Annual neurologic evaluation to identify potential complications of spasticity, such as contractures.

Genetic counseling

SPG7 is inherited in an autosomal recessive manner. Heterozygotes (carriers) are usually asymptomatic. Each sib of an affected individual has a 25% chance of being affected, a 50% chance of being an asymptomatic carrier, and a 25% chance of being unaffected and not a carrier. Carrier testing for at-risk relatives, prenatal testing for a

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pregnancy at increased risk and preimplantation genetic testing are possible if both pathogenic alleles have been identified in the family.

Diagnosis

Suggestive Findings

Spastic paraplegia 7 (SPG7) should be suspected in individuals with the following:

- Insidiously progressive bilateral leg weakness
- Spasticity
- Decreased vibratory sense
- Cerebellar signs
- Neurologic examination demonstrating EITHER of the following:
 - A pure phenotype of spastic paraplegia with hyperreflexia, extensor plantar responses, and mildly impaired vibration sensation in the distal legs
 - A complicated phenotype of spastic paraplegia including optic neuropathy, progressive external ophthalmoplegia/ptosis slowed speech, swallowing difficulties, palatal tremor, subtle cognitive impairment, urinary urgency, ataxia, nystagmus, strabismus, decreased hearing, scoliosis, *pes cavus*, motor and sensory neuropathy, and amyotrophy [Brugman et al 2008, Salinas et al 2008, Warnecke et al 2010, Pfeffer et al 2014]
- Neuroimaging findings of cerebellar atrophy (MRI) or white matter changes as detected by diffusion tensor imaging in the frontal lobes, the corticospinal tracts, and the brain stem
- Family history consistent with autosomal recessive inheritance

Establishing the Diagnosis

The diagnosis of SPG 7 **is established** in a proband with typical clinical findings and identification of biallelic pathogenic variants in *SPG7* by molecular genetic testing (see Table 1).

Note: A single *SPG7* pathogenic variant (p.Leu78^{*}) was identified in a proband with a pure HSP phenotype suggesting that heterozygosity for an *SPG7* pathogenic variant may be sufficient to cause disease (i.e., autosomal dominant inheritance). However, this conclusion is challenged by the finding of unaffected p.Leu78^{*} heterozygotes in other families as well as the possibility that the affected heterozygous individual had a second *SPG7* pathogenic variant which was not detected due to testing limitations [Sánchez-Ferrero et al 2013].

Because the phenotype of SPG7 is indistinguishable from many other forms of hereditary spastic paraplegia, recommended molecular genetic testing approaches include use of a **multigene panel** or **comprehensive genomic testing**.

Note: Single-gene testing (sequence analysis of *SPG7*, followed by gene-targeted deletion/duplication analysis) is rarely useful and typically NOT recommended.

• A multigene panel that includes *SPG7* and other genes of interest (see Differential Diagnosis) is most likely to identify the genetic cause of the condition at the most reasonable cost while limiting identification of variants of uncertain significance and pathogenic variants in genes that do not explain the underlying phenotype. Note: (1) The genes included in the panel and the diagnostic sensitivity of the testing used for each gene vary by laboratory and are likely to change over time. (2) Some multigene panels may include genes not associated with the condition discussed in this *GeneReview*. (3) In some laboratories, panel options may include a custom laboratory-designed panel and/or custom phenotype-focused exome analysis that includes genes specified by the clinician. (4) Methods used in a panel may include sequence

analysis, deletion/duplication analysis, and/or other non-sequencing-based tests. For this disorder a multigene panel that also includes deletion/duplication analysis is recommended (see Table 1).

For an introduction to multigene panels click here. More detailed information for clinicians ordering genetic tests can be found here.

• **Comprehensive genomic testing** (which does not require the clinician to determine which gene[s] are likely involved) is another good option. **Exome sequencing** is most commonly used; **genome sequencing** is also possible. Exome array (when clinically available) may be considered if exome sequencing is not diagnostic.

For an introduction to comprehensive genomic testing click here. More detailed information for clinicians ordering genomic testing can be found here.

Gene ¹	Method	Proportion of Pathogenic Variants ² Detectable by Method		
	Sequence analysis ³	>98% ⁴		
SPG7	Gene-targeted deletion/duplication analysis ⁵	<2% ⁶		

Table 1. Molecular Genetic Testing Used in Spastic Paraplegia 7

1. See Table A. Genes and Databases for chromosome locus and protein.

2. See Molecular Genetics for information on allelic variants detected in this gene.

3. Sequence analysis detects variants that are benign, likely benign, of uncertain significance, likely pathogenic, or pathogenic. Variants may include small intragenic deletions/insertions and missense, nonsense, and splice site variants; typically, exon or whole-gene deletions/duplications are not detected. For issues to consider in interpretation of sequence analysis results, click here.

4. Arnoldi et al [2008], Brugman et al [2008], Klebe et al [2012], van Gassen et al [2012], Sánchez-Ferrero et al [2013], Pfeffer et al [2015]

5. Gene-targeted deletion/duplication analysis detects intragenic deletions or duplications. Methods used may include quantitative PCR, long-range PCR, multiplex ligation-dependent probe amplification (MLPA), and a gene-targeted microarray designed to detect single-exon deletions or duplications.

6. Casari et al [1998], Arnoldi et al [2008], Klebe et al [2012], van Gassen et al [2012], Sánchez-Ferrero et al [2013]

Clinical Characteristics

Clinical Description

Spastic paraplegia 7 (SPG7) is characterized by insidiously progressive bilateral lower-limb weakness and spasticity. Most affected individuals have proximal or generalized weakness in the legs and impaired vibration sense.

Onset typically occurs in adulthood, around age 30-45 years, although symptoms may start as early as age 11 years and as late as age 72 years [De Michele et al 1998, McDermott et al 2001, Wilkinson et al 2004].

Presentation. The first sign is typically insidiously progressive bilateral leg weakness.

Additional features. Other signs and symptoms can be observed [Brugman et al 2008, Salinas et al 2008, Warnecke et al 2010, Almontashiri et al 2014, Pfeffer et al 2014] including the following:

- Cerebellar and motor signs
 - Ataxia (gait and limbs)
 - Spastic dysarthria
 - Dysphagia
- Ophthalmic findings
 - Pale optic disks

- Nystagmus
- Strabismus
- Ptosis
- Hearing loss of conductive/neurosensory /mixed type
- Peripheral neuromuscular findings
 - Motor and sensory neuropathy
 - Amyotrophy
- Orthopedic issues
 - Scoliosis
 - Pes cavus
- Urinary sphincter disturbances

Progression. Severe disability of gait due to leg spasticity may develop as soon as eight years after onset of symptoms, and some individuals are confined to a wheelchair [Elleuch et al 2006, Schüle et al 2006].

Findings on Neuroimaging and Other Investigations

Neuroimaging

- In a few individuals, conventional cerebral MRI may show cerebellar (or, less frequently, cortical) atrophy [Salinas et al 2008, Hourani et al 2009, Warnecke et al 2010].
- White matter changes as detected by diffusion tensor imaging in the frontal lobes, the corticospinal tracts, and the brain stem are specific to SPG7.
- Spinal imaging studies are useful in the differential diagnosis to exclude other anomalies of the pontomedullary junction and of the cervical and dorsolumbar medulla.

Other investigations

- Spinal evoked potentials may reveal delayed prolongation of the central conduction time [Nielsen et al 2001].
- Electromyography with nerve conduction velocities may reveal axonal sensory motor neuropathy.
- Paired transcranial magnetic stimulation may show delayed prolongation of the central motor conduction time and motor threshold in some affected individuals in lower limb muscles [Warnecke et al 2010]. Intracortical inhibition appears normal in SPG7 [Nardone & Tezzon 2003].
- Optical coherence tomography is useful for detecting subclinical optic neuropathy [Klebe et al 2012].
- A battery of neuropsychological tests may reveal mild impairment of visuoconstructive and executive functions in some individuals [Warnecke et al 2010].
- Serum creatine kinase activity may be slightly above the normal range in some cases.
- Muscle biopsy has revealed the following:
 - Changes of denervation with partial reinnervation
 - Atrophic, angulated fibers, predominantly type II
 - Ragged-red fibers, which are positive for the histoenzymatic reaction to succinate dehydrogenase and negative for cytochrome *c* oxidase (COX, the complex IV of the mitochondrial respiratory chain), indicating an oxidative phosphorylation defect [Casari et al 1998, McDermott et al 2001, Wilkinson et al 2004, Tzoulis et al 2008].

Genotype-Phenotype Correlations

No genotype-phenotype correlations can be proposed based on published studies.

Prevalence

The prevalence of SPG7 is estimated at between 1:100,000 and 9:100,000 for most countries (www.orpha.net).

Genetically Related (Allelic) Disorders

Primary lateral sclerosis (PLS). Compound heterozygous pathogenic variants in *SPG7* were reported as cosegregating with PLS in an autosomal recessive manner in five individuals from the same family [Yang et al 2016].

Amyotrophic lateral sclerosis (ALS). Four individuals with nonfamilial ALS were found to heterozygous pathogenic variants in *SPG7* [Krüger et al 2016].

Spinocerebellar ataxia (SCA). It is often difficult to clinically discriminate between hereditary spastic paraplegia (HSP) and SCA as the disorders can share multiple features. Several *SPG7* pathogenic variants have been reported to be associated with an ataxia phenotype [Pfeffer et al 2015, Choquet et al 2016, Synofzik & Schule 2017].

Differential Diagnosis

No significant differences exist between spastic paraplegia 7 (SPG7) and other types of pure autosomal dominant and autosomal recessive spastic paraplegia [Fink 2002, Fink 2003, Salinas et al 2008] (see Hereditary Spastic Paraplegia Overview for a review). However, Brugman et al [2008] reported that *SPG7* pathogenic variants are less likely to be found in adult-onset cases in which upper motor neuron symptoms (UMN) are present in the arms and in adult-onset cases with UMN symptoms involving the bulbar region.

Other conditions that need to be considered in the differential diagnosis of SPG7 are summarized in Table 2.

Table 2. Other Disorders to Consider in the Differential Diagnosis of SPG7

DiffDx Disorder	Gene(s)	MOI	Clinical Features of the DiffDx Disorder		
DinDx Disorder	Gene(s)		Overlapping w/SPG7	Distinguishing from SPG7	
Adrenomyeloneuropathy and other leukodystrophies (e.g., Krabbe disease, arylsulfatase A deficiency [metachromatic leukodystrophy])	ABCD1 GALC ARSA	XL AR	Paraplegia neuropathy	 Dementia On MRI: leukodystrophy, adrenal dysfunction, long-chain fatty acid accumulation 	
Spinocerebellar ataxia type 28	AFG3L2	AD	Paraplegia; ataxia	Rare dystonia or parkinsonism	
Dopa-responsive dystonia	GCH1	AD	Brisk reflexes; spasticity; extensor plantar responses	Young-onset dystonia parkinsonism responsive to levodopaDiurnal variation	
Amyotrophic lateral sclerosis	See footnote 1.	AD AR XL	Spasticity	Muscle atrophy, weakness & fasciculations	
Primary lateral sclerosis ²	Unknown	NA	Spasticity	Survival 15-20 years	
Arginase deficiency	ARG1	AR	Spasticity	 Epileptic seizures Severe mental retardation ↑ plasma arginine Hyperammonemia 	
Structural abnormalities of the brain or spinal cord	NA	NA	Gait difficulties	On MRI: spine abnormalities	

Table 2. continued from previous page.

DiffDx Disorder	Gene(s)	MOI	Clinical Features of the DiffDx Disorder	
DiiDx Disoidei	Gene(s)	WOI	Overlapping w/SPG7	Distinguishing from SPG7
Vitamin B ₁₂ deficiency	NA	NA	Unsteady gait	 Subacute combined degeneration Improvement after vitamin B₁₂ supplementation
Primary progressive multiple sclerosis	NA	NA	Spasticity	 MRI white matter changes Oligoclonal IgG bands ↑ IgG index
Progressive external ophthalmoplegia	Various	AR AD	Eyelid ptosis	External ophthalmoplegiaProximal myopathyNo pyramidal signs
Tropical spastic paraplegia (caused by HTLV1 infection)	NA	NA	Paraplegia	HTLV-1 serology
Optic neuropathy	KLC2 ³ MFN2 ⁴	AR AD	Pale optic disks	No pyramidal signs

AD = autosomal dominant; AR = autosomal recessive; DiffDx = differential diagnosis; MOI = mode of inheritance; NA = not applicable; XL = X-linked

1. See Phenotypic Series: Amyotrophic lateral sclerosis for a list of genes associated with this phenotype in OMIM.

- 2. Brugman et al [2008]
- 3. Melo et al [2015]

4. Züchner et al [2006]

Management

Evaluations Following Initial Diagnosis

To establish the extent of disease and needs in an individual diagnosed with spastic paraplegia 7 (SPG7), the following evaluations are recommended if they have not already been completed:

- Ophthalmologic evaluation
- Hearing testing
- Urologic evaluation in case of bladder dysfunction
- Consultation with a clinical geneticist and/or genetic counselor

Evaluation by a multidisciplinary team that includes a general practitioner, neurologist, physical therapist, social worker, and psychologist should be considered.

Neuropsychological testing may be suggested.

Treatment of Manifestations

No specific drug treatments or cures for SPG7 exist.

Drugs to reduce spasticity and muscle tightness include baclofen, tizanidine, dantrolene, and diazepam – preferably administered one at a time.

Management of spasticity by intrathecal baclofen or intramuscular botulinum toxin injections may be an option in selected individuals [Kawano et al 2018].

A combination of physical therapy and assistive walking devices are often used to reduce contractures, provide support, and promote stability.

Occupational therapy and speech therapy are often helpful in managing activities of daily living.

Prevention of Secondary Complications

Because individuals with advanced disease are bedridden they are at major risk of aspiration pneumonia, urinary tract infections and pulmonary embolism; careful monitoring is recommended to help avoid these complications.

Surveillance

Annual neurologic evaluation can help identify potential complications of spasticity that develop over time (e.g., contractures).

Evaluation of Relatives at Risk

See Genetic Counseling for issues related to testing of at-risk relatives for genetic counseling purposes.

Therapies Under Investigation

Search ClinicalTrials.gov in the US and EU Clinical Trials Register in Europe for access to information on clinical studies for a wide range of diseases and conditions. Note: There may not be clinical trials for this disorder.

Genetic Counseling

Genetic counseling is the process of providing individuals and families with information on the nature, mode(s) of inheritance, and implications of genetic disorders to help them make informed medical and personal decisions. The following section deals with genetic risk assessment and the use of family history and genetic testing to clarify genetic status for family members; it is not meant to address all personal, cultural, or ethical issues that may arise or to substitute for consultation with a genetics professional. —ED.

Mode of Inheritance

Spastic paraplegia 7 (SPG7) is inherited in an autosomal recessive manner.

Risk to Family Members

Parents of a proband

- The parents of an affected individual are obligate heterozygotes (i.e., carriers of one *SPG7* pathogenic variant).
- Findings of a subtle reduction of white matter integrity in the corpus callosum on diffusion tensor imaging have been reported in individuals heterozygous for an *SPG7* pathogenic variant [Warnecke et al 2010] and a single possibly manifesting heterozygote has been reported [Sánchez-Ferrero et al 2013]. However, heterozygotes are typically not at risk of developing the disorder.

Sibs of a proband

- At conception, each sib of an affected individual has a 25% chance of being affected, a 50% chance of being an asymptomatic carrier, and a 25% chance of being unaffected and not a carrier.
- Heterozygotes (carriers) are asymptomatic and typically are not at risk of developing the disorder (see **Parents of a proband**).

Offspring of a proband. The offspring of an individual with SPG7 are obligate heterozygotes (carriers) for a pathogenic variant in *SPG7*.

Other family members. Each sib of the proband's parents is at a 50% risk of being a carrier of a *SPG7* pathogenic variant.

Carrier (Heterozygote) Detection

Carrier testing for at-risk relatives requires prior identification of the SPG7 pathogenic variants in the family.

Related Genetic Counseling Issues

Family planning

- The optimal time for determination of genetic risk, clarification of carrier status, and discussion of the availability of prenatal/preimplantation genetic testing is before pregnancy.
- It is appropriate to offer genetic counseling (including discussion of potential risks to offspring and reproductive options) to young adults who are affected, are carriers, or are at risk of being carriers.

DNA banking is the storage of DNA (typically extracted from white blood cells) for possible future use. Because it is likely that testing methodology and our understanding of genes, allelic variants, and diseases will improve in the future, consideration should be given to banking DNA of affected individuals.

Prenatal Testing and Preimplantation Genetic Testing

Once the *SPG7* pathogenic variants have been identified in an affected family member, prenatal testing for a pregnancy at increased risk and preimplantation genetic testing for SPG7 are possible.

Differences in perspective may exist among medical professionals and within families regarding the use of prenatal testing. While most centers would consider use of prenatal testing to be a personal decision, discussion of these issues may be helpful.

Resources

GeneReviews staff has selected the following disease-specific and/or umbrella support organizations and/or registries for the benefit of individuals with this disorder and their families. GeneReviews is not responsible for the information provided by other organizations. For information on selection criteria, click here.

HSP Research Foundation

Australia Email: inquiries@hspersunite.org.au www.hspersunite.org.au

• National Institute of Neurological Disorders and Stroke (NINDS)

Phone: 800-352-9424 Hereditary Spastic Paraplegia Information Page

 Spastic Paraplegia Foundation, Inc. Phone: 877-773-4483 sp-foundation.org

• Tom Wahlig-Foundation

Tom Wahlig Stiftung Germany www.hsp-info.de/en/foundation.htm

• A.I. Vi.P.S.

Associazione Italiana Vivere la Paraparesi Spastica Via Tevere, 7 20020 Lainate (MI) Italy Phone: 39 392 9825622 Email: info@aivips.it www.aivips.it

Molecular Genetics

Information in the Molecular Genetics and OMIM tables may differ from that elsewhere in the GeneReview: tables may contain more recent information. —ED.

Table A. Spastic Paraplegia 7: Genes and Databases

Gene	Chromosome Locus	Protein	Locus-Specific Databases	HGMD	ClinVar
SPG7	16q24.3	Paraplegin	alsod/SPG7 genetic mutations SPG7 database	SPG7	SPG7

Data are compiled from the following standard references: gene from HGNC; chromosome locus from OMIM; protein from UniProt. For a description of databases (Locus Specific, HGMD, ClinVar) to which links are provided, click here.

Table B. OMIM Entries for Spastic Paraplegia 7 (View All in OMIM)

602783 SPG7 MATRIX AAA PEPTIDASE SUBUNIT, PARAPLEGIN; SPG7

607259 SPASTIC PARAPLEGIA 7, AUTOSOMAL RECESSIVE; SPG7

Gene structure. *SPG7* spans a physical distance of approximately 52 kb and comprises 17 exons. See Table A, **Gene** for a detailed summary of gene and protein information.

Pathogenic variants. Pathogenic missense, nonsense, frameshift, and splice site variants have been observed throughout *SPG7*. Missense variants occur most frequently. Missense and truncating variants, such as c.1454_1462del9 (reported as c.1450_1458del9 [McDermott et al 2001]), deletion of 9.5 kb [Casari et al 1998] and 2228insA [Casari et al 1998] have been reported to delete main protein functional domains.

Twenty-seven pathogenic variants have been reported in *SPG7* [Casari et al 1998, Arnoldi et al 2008, Brugman et al 2008, Klebe et al 2012, van Gassen et al 2012, Sánchez-Ferrero et al 2013, Pfeffer et al 2015]. The *SPG7* c.1529C>T (p.Ala510Val) variant is the most frequent variant found across populations [Sánchez-Ferrero et al 2013, Pfeffer et al 2015, Choquet et al 2016]. Although this variant and most others identified have been associated with an ataxic phenotype, recent efforts have focused on associating *SPG7* variants with additional clinical features. Recent studies suggest that cerebellar ataxia is a frequent feature among individuals with *SPG7*-related disease [Pfeffer et al 2015, Synofzik & Schule 2017]. The ataxic syndrome could even be a predominant feature over spasticity, as observed in a Japanese family segregating p.Arg398Ter [Yahikozawa et al 2015].

DNA Nucleotide Change (Alias) ¹	Predicted Protein Change	Reference Sequences
del9.5 kb		
c.1053dupC		
c.1192C>T	p.Arg398Ter	
c.1454_1462del9	p.Arg485_Glu487del	NM_003119.3
c.1529C>T	p.Ala510Val	NP_003110.1
c.2102A>C	p.His701Pro	
c.2216insA (2228insA ²)		

Table 3. SPG7 Pathogenic Variants Discussed in This GeneReview

Variants listed in the table have been provided by the authors. *GeneReviews* staff have not independently verified the classification of variants.

GeneReviews follows the standard naming conventions of the Human Genome Variation Society (varnomen.hgvs.org). See Quick Reference for an explanation of nomenclature.

1. Variant designation that does not conform to current naming conventions

2. Casari et al [1998]

Normal gene product. Paraplegin is a mitochondrial inner membrane protein that exerts protein quality control in a high molecular complex with AFG3L2. Both paraplegin and AFG3L2 belong to the AAA protein family (ATPases associated with diverse cellular activities) (see also Hereditary Spastic Paraplegia Overview). Paraplegin and AFG3L2 coassemble in the mitochondrial inner membrane, forming a high molecular-weight complex [Atorino et al 2003]. Paraplegin is ubiquitously expressed in adult and fetal human tissues. The presence of two hydrophobic regions, which have the characteristics of transmembrane domains, allows identification of both paraplegin and AFG3L2 as integral membrane proteins. The AAA domain is in the central part of paraplegin between amino acid residues 344 and 534, while a coil-coil domain in the carboxy-terminal part of the molecule promotes assembly in the hexameric complex. In order to achieve maturation, paraplegin undergoes several cleavages upon its import in the mitochondria inner membrane by the mitochondrial processing peptidase and by the m-AAA protease complex itself [Koppen et al 2009]. Thus, the final processing of paraplegin in a mature form depends on its coassembly with AFG3L2 and, as recently demonstrated, the unphosphorylation of AFG3L2 in position p.Tyr179 [Almontashiri et al 2014].

In a recent paper paraplegin was identified as a molecular component/regulator of the mitochondrial permeability transition pore [Shanmughapriya et al 2015]; however another study demonstrated conflicting results [König et al 2016].

Abnormal gene product. Inactivation of the paraplegin-AFG3L2 complex causes reduced complex I activity in mitochondria. Loss of AFG3L2 function is associated with autosomal recessive spastic ataxia 5 and spinocerebellar ataxia 28 (see Differential Diagnosis).

Biochemical analysis from two individuals with confirmed *SPG7* pathogenic variants revealed a reduction in citrate synthase-corrected complex I and complex II/III activities in muscle and complex I activity in mitochondrial-enriched fractions from cultured myoblasts. Mitochondrial DNA damage has been observed in muscle biopsies of affected individuals with *SPG7* variants c.2102A>C and c.1053dupC [Tzoulis et al 2008, Wedding et al 2014]. Further studies should clarify how paraplegin can alter mitochondrial DNA; however, this could be an indirect effect of the dysfunction of the mAAA-protease complex, as paraplegin does not interact directly with the DNA.

In mouse, *AFG3L2* homozygous pathogenic variants appear more severe than paraplegin variants; null or missense *Afg3l2* mouse models developed marked impairment of axonal development leading to neonatal death

[Maltecca et al 2008]. The mice developed a severe early-onset tetraparesis and were found to have reduced myelinated fibers in the spinal cord and impaired respiratory chain complex I and III activity. The increased severity of the phenotype is explained by the higher neuronal expression of AFG3L2, but also the ability of AFG3L2 to form homocomplexes, while paraplegin requires coassembly with AFG3L2 to form functional complexes. Heterozygous pathogenic variants of *AFG3L2* have been associated with a dominant form of spinocerebellar ataxia (SCA28) [Di Bella et al 2010].

Gene expression regulation. *SPG7* is one of the targets of miR-224, which is located in an intron of the GABA A receptor ε subunit (GABRE) and produced in several models of cancer proliferation [Fu et al 2016].

References

Literature Cited

- Almontashiri NA, Chen HH, Mailloux RJ, Tatsuta T, Teng AC, Mahmoud AB, Ho T, Stewart NA, Rippstein P, Harper ME, Roberts R, Willenborg C, Erdmann J, Pastore A, McBride HM, Langer T, Stewart AF, et al. SPG7 variant escapes phosphorylation-regulated processing by AFG3L2, elevates mitochondrial ROS, and is associated with multiple clinical phenotypes. Cell Rep. 2014;7:834–47. PubMed PMID: 24767997.
- Arnoldi A, Tonelli A, Crippa F, Villani G, Pacelli C, Sironi M, Pozzoli U, D'Angelo MG, Meola G, Martinuzzi A, Crimella C, Redaelli F, Panzeri C, Renieri A, Comi GP, Turconi AC, Bresolin N, Bassi MT. A clinical, genetic, and biochemical characterization of SPG7 mutations in a large cohort of patients with hereditary spastic paraplegia. Hum Mutat. 2008;29:522–31. PubMed PMID: 18200586.
- Atorino L, Silvestri L, Koppen M, Cassina L, Ballabio A, Marconi R, Langer T, Casari G. Loss of m-AAA protease in mitochondria causes complex I deficiency and increased sensitivity to oxidative stress in hereditary spastic paraplegia. J Cell Biol. 2003;163:777–87. PubMed PMID: 14623864.
- Brugman F, Scheffer H, Wokke JH, Nillesen WM, de Visser M, Aronica E, Veldink JH, van den Berg LH. Paraplegin mutations in sporadic adult-onset upper motor neuron syndromes. Neurology. 2008;71:1500–5. PubMed PMID: 18799786.
- Casari G, De Fusco M, Ciarmatori S, Zeviani M, Mora M, Fernandez P, De Michele G, Filla A, Cocozza S, Marconi R, Dürr A, Fontaine B, Ballabio A. Spastic paraplegia and OXPHOS impairment caused by mutations in paraplegin, a nuclear-encoded mitochondrial metalloprotease. Cell. 1998;93:973–83. PubMed PMID: 9635427.
- Choquet K, Tetreault M, Yang S, La Piana R, Dicaire MJ, Vanstone MR, Mathieu J, Bouchard JP, Rioux MF, Rouleau GA, Boycott KM, Majewski J, Brais B, et al. SPG7 mutations explain a significant proportion of French Canadian spastic ataxia cases. Eur J Hum Genet. 2016;24:1016–21. PubMed PMID: 26626314.
- De Michele G, De Fusco M, Cavalcanti F, Filla A, Marconi R, Volpe G, Monticelli A, Ballabio A, Casari G, Cocozza S. A new locus for autosomal recessive hereditary spastic paraplegia maps to chromosome 16q24.3. Am J Hum Genet. 1998;63:135–9. PubMed PMID: 9634528.
- Di Bella D, Lazzaro F, Brusco A, Plumari M, Battaglia G, Pastore A, Finardi A, Cagnoli C, Tempia F, Frontali M, Veneziano L, Sacco T, Boda E, Brussino A, Bonn F, Castellotti B, Baratta S, Mariotti C, Gellera C, Fracasso V, Magri S, Langer T, Plevani P, Di Donato S, Muzi-Falconi M, Taroni F. Mutations in the mitochondrial protease gene AFG3L2 cause dominant hereditary ataxia SCA28. Nature Genetics. 2010;42:313–21. PubMed PMID: 20208537.
- Elleuch N, Depienne C, Benomar A, Hernandez AM, Ferrer X, Fontaine B, Grid D, Tallaksen CM, Zemmouri R, Stevanin G, Durr A, Brice A. Mutation analysis of the paraplegin gene (SPG7) in patients with hereditary spastic paraplegia. Neurology. 2006;66:654–9. PubMed PMID: 16534102.
- Fink JK. Hereditary spastic paraplegia. Neurol Clin. 2002;20:711-26. PubMed PMID: 12432827.

- Fink JK. The hereditary spastic paraplegias: nine genes and counting. Arch Neurol. 2003;60:1045–9. PubMed PMID: 12925358.
- Fu F, Wu D, Qian C. The MicroRNA-224 inhibitor prevents neuronal apoptosis via targeting spastic paraplegia 7 after cerebral ischemia. J Mol Neurosci. 2016;59:421–9. PubMed PMID: 27165196.
- Hourani R, El-Hajj T, Barada WH, Hourani M, Yamout BI. MR imaging findings in autosomal recessive hereditary spastic paraplegia. AJNR Am J Neuroradiol. 2009;30:936–40. PubMed PMID: 19193756.
- Kawano O, Masuda M, Takao T, Sakai H, Morishita Y, Hayashi T, Ueta T, Maeda T. The dosage and administration of long-term intrathecal baclofen therapy for severe spasticity of spinal origin. Spinal Cord. 2018;56:996–9. PubMed PMID: 29895878.
- Klebe S, Depienne C, Gerber S, Challe G, Anheim M, Charles P, Fedirko E, Lejeune E, Cottineau J, Brusco A, Dollfus H, Chinnery PF, Mancini C, Ferrer X, Sole G, Destée A, Mayer JM, Fontaine B, de Seze J, Clanet M, Ollagnon E, Busson P, Cazeneuve C, Stevanin G, Kaplan J, Rozet JM, Brice A, Durr A. Spastic paraplegia gene 7 in patients with spasticity and/or optic neuropathy. Brain. 2012;135:2980–93. PubMed PMID: 23065789.
- König T, Tröder SE, Bakka K, Korwitz A, Richter-Dennerlein R, Lampe PA, Patron M, Mühlmeister M, Guerrero-Castillo S, Brandt U, Decker T, Lauria I, Paggio A, Rizzuto R, Rugarli EI, De Stefani D, Langer T. The m-AAA protease associated with neurodegeneration limits MCU activity in mitochondria. Mol Cell. 2016;64:148–62. PubMed PMID: 27642048.
- Koppen M, Bonn F, Ehses S, Langer T. Autocatalytic processing of m-AAA protease subunits in mitochondria. Mol Biol Cell. 2009;20:4216–24. PubMed PMID: 19656850.
- Krüger S, Battke F, Sprecher A, Munz M, Synofzik M, Schols L, Gasser T, Grehl T, Prudlo J, Biskup S. Rare variants in neurodegeneration associated genes revealed by targeted panel sequencing in a German ALS cohort. Front Mol Neurosci. 2016;9:92. PubMed PMID: 27790088.
- Maltecca F, Aghaie A, Schroeder DG, Cassina L, Taylor BA, Phillips SJ, Malaguti M, Previtali S, Guénet JL, Quattrini A, Cox GA, Casari G. The mitochondrial protease AFG3L2 is essential for axonal development. J Neurosci. 2008;28:2827–36. PubMed PMID: 18337413.
- McDermott CJ, Dayaratne RK, Tomkins J, Lusher ME, Lindsey JC, Johnson MA, Casari G, Turnbull DM, Bushby K, Shaw PJ. Paraplegin gene analysis in hereditary spastic paraparesis (HSP) pedigrees in northeast England. Neurology. 2001;56:467–71. PubMed PMID: 11222789.
- Melo US, Macedo-Souza LI, Figueiredo T, Muotri AR, Gleeson JG, Coux G, Armas P, Calcaterra NB, Kitajima JP, Amorim S, Olavio TR, Griesi-Oliveira K, Coatti GC, Rocha CRR, Martins-Pinheiro M, Menck CFM, Zaki MS, Kok F, Zatz M, Santos S. Overexpression of KLC2 due to a homozygous deletion in the non-coding region causes SPOAN syndrome. Hum Mol Genet. 2015;24:6877–85. PubMed PMID: 26385635.
- Nardone R, Tezzon F. Transcranial magnetic stimulation study in hereditary spastic paraparesis. Eur Neurol. 2003;49:234–7. PubMed PMID: 12736541.
- Nielsen JE, Jennum P, Fenger K, Sørensen SA, Fuglsang-Frederiksen A. Increased intracortical facilitation in patients with autosomal dominant pure spastic paraplegia linked to chromosome 2p. Eur J Neurol. 2001;8:335. PubMed PMID: 11422430.
- Pfeffer G, Gorman GS, Griffin H, Kurzawa-Akanbi M, Blakely EL, Wilson I, Sitarz K, Moore D, Murphy JL, Alston CL, Pyle A, Coxhead J, Payne B, Gorrie GH, Longman C, Hadjivassiliou M, McConville J, Dick D, Imam I, Hilton D, Norwood F, Baker MR, Jaiser SR, Yu-Wai-Man P, Farrell M, McCarthy A, Lynch T, McFarland R, Schaefer AM, Turnbull DM, Horvath R, Taylor RW, Chinnery PF. Mutations in the SPG7 gene cause chronic progressive external ophthalmoplegia through disordered mitochondrial DNA maintenance. Brain. 2014;137:1323–36. PubMed PMID: 24727571.

- Pfeffer G, Pyle A, Griffin H, Miller J, Wilson V, Turnbull L, Fawcett K, Sims D, Eglon G, Hadjivassiliou M, Horvath R, Németh A, Chinnery PF. SPG7 mutations are a common cause of undiagnosed ataxia. Neurology. 2015;84:1174–6. PubMed PMID: 25681447.
- Salinas S, Proukakis C, Crosby A, Warner TT. Hereditary spastic paraplegia: clinical features and pathogenetic mechanisms. Lancet Neurol. 2008;7:1127–38. PubMed PMID: 19007737.
- Sánchez-Ferrero E, Coto E, Beetz C, Gámez J, Corao AI, Díaz M, Esteban J, del Castillo E, Moris G, Infante J, Menéndez M, Pascual-Pascual SI, López de Munaín A, Garcia-Barcina MJ, Alvarez V, et al. SPG7 mutational screening in spastic paraplegia patients supports a dominant effect for some mutations and a pathogenic role for p.A510V. Clin Genet. 2013;83:257–62. PubMed PMID: 22571692.
- Schüle R, Holland-Letz T, Klimpe S, Kassubek J, Klopstock T, Mall V, Otto S, Winner B, Schöls L. The Spastic Paraplegia Rating Scale (SPRS): a reliable and valid measure of disease severity. Neurology. 2006;67:430–4. PubMed PMID: 16894103.
- Shanmughapriya S, Rajan S, Hoffman NE, Higgins AM, Tomar D, Nemani N, Hines KJ, Smith DJ, Eguchi A, Vallem S, Shaikh F, Cheung M, Leonard NJ, Stolakis RS, Wolfers MP, Ibetti J, Chuprun JK, Jog NR, Houser SR, Koch WJ, Elrod JW, Madesh M. SPG7 is an essential and conserved component of the mitochondrial permeability transition pore. Mol Cell. 2015;60:47–62. PubMed PMID: 26387735.
- Synofzik M, Schule R. Overcoming the divide between ataxias and spastic paraplegias: Shared phenotypes, genes, and pathways. Mov Disord. 2017;32:332–45. PubMed PMID: 28195350.
- Tzoulis C, Denora PS, Santorelli FM, Bindoff LA. Hereditary spastic paraplegia caused by the novel mutation 1047insC in the SPG7 gene. J Neurol. 2008;255:1142–4. PubMed PMID: 18563470.
- van Gassen KL, van der Heijden CD, de Bot ST, den Dunnen WF, van den Berg LH, Verschuuren-Bemelmans CC, Kremer HP, Veldink JH, Kamsteeg EJ, Scheffer H, van de Warrenburg BP. Genotype-phenotype correlations in spastic paraplegia type 7: a study in a large Dutch cohort. Brain. 2012;135:2994–3004. PubMed PMID: 22964162.
- Warnecke T, Duning T, Schirmacher A, Mohammadi S, Schwindt W, Lohmann H, Dziewas R, Deppe M, Ringelstein EB, Young P. A novel splice site mutation in the SPG7 gene causing widespread fiber damage in homozygous and heterozygous subjects. Mov Disord. 2010;25:413–20. PubMed PMID: 20108356.
- Wedding IM, Koht J, Tran GT, Misceo D, Selmer KK, Holmgren A, Frengen E, Bindoff L, Tallaksen CM, Tzoulis C. Spastic paraplegia type 7 is associated with multiple mitochondrial DNA deletions. PLoS One. 2014;9:e86340. PubMed PMID: 24466038.
- Wilkinson PA, Crosby AH, Turner C, Bradley LJ, Ginsberg L, Wood NW, Schapira AH, Warner TT. A clinical, genetic and biochemical study of SPG7 mutations in hereditary spastic paraplegia. Brain. 2004;127:973–80. PubMed PMID: 14985266.
- Yahikozawa H, Yoshida K, Sato S, Hanyu N, Doi H, Miyatake S, Matsumoto N. Predominant cerebellar phenotype in spastic paraplegia 7 (SPG7). Hum Genome Var. 2015;2:15012. PubMed PMID: 27081526.
- Yang Y, Zhang L, Lynch DR, Lukas T, Ahmeti K, Sleiman PM, Ryan E, Schadt KA, Newman JH, Deng HX, Siddique N, Siddique T. Compound heterozygote mutations in SPG7 in a family with adult-onset primary lateral sclerosis. Neurol Genet. 2016;2:e60. PubMed PMID: 27123479.
- Züchner S, De Jonghe P, Jordanova A, Claeys KG, Guergueltcheva V, Cherninkova S, Hamilton SR, Van Stavern G, Krajewski KM, Stajich J, Tournev I, Verhoeven K, Langerhorst CT, de Visser M, Baas F, Bird T, Timmerman V, Shy M, Vance JM. Axonal neuropathy with optic atrophy is caused by mutations in mitofusin 2. Ann Neurol. 2006;59:276–81. PubMed PMID: 16437557.

Chapter Notes

Revision History

- 25 October 2018 (ha) Comprehensive update posted live
- 23 December 2010 (me) Comprehensive update posted live
- 25 February 2008 (cd) Revision: deletion/duplication analysis available clinically
- 24 August 2006 (me) Review posted live
- 7 March 2005 (gc) Original submission

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